SYNTHESIS OF ARACHIDONIC ACID METABOLITES PRODUCED BY PURIFIED KIDNEY CORTEX MICROSOMAL CYTOCHROME P-450

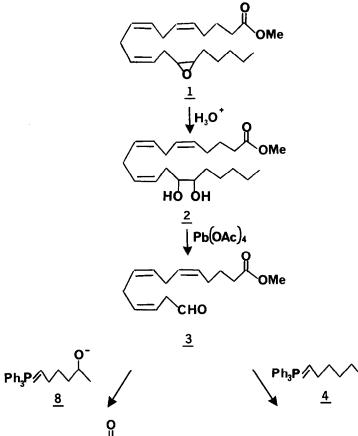
> Sukumar Manna, J.R. Falck^{*} Department of Molecular Genetics

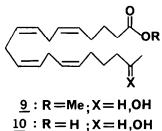
N. Chacos and J. Capdevila Department of Biochemistry University of Texas Health Science Center at Dallas Dallas, Texas 75235 USA

<u>Summary</u>: Reported are the synthesis and structure confirmation of the major metabolites produced during the NADPH dependent oxidation of arachidonic acid by a reconstituted enzyme system containing purified kidney cortex microsomal cytochrome P-450.

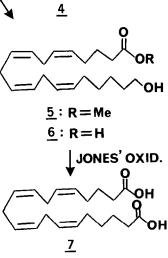
Recently, Capdevila¹ and Others² have demonstrated a NADPH-dependent cytochrome P-450 mediated pathway for the rapid and efficient conversion of arachidonic acid to a rich variety of oxygenated metabolites. In addition to various lipoxygenase-type products, several novel metabolites unique to the cytochrome pathway have been isolated³, some of which have potent and meaningful biological activity⁴. Significantly, both the identity and ratio of metabolites are associated closely with the tissue source from which the cytochrome P-450 is isolated^{1c}. In the case of cytochrome P-450 purified from pig kidney cortex microsomes, the major products⁵ of incubation with arachidonic acid are 19- and 20-hydroxyeicosatetraenoic acid, 19-oxoeicosatetraenoic acid, and eicosatetraen-1,20-dioic acid, (10), (6), (11), and (7), respectively. These metabolites have been reported previously² from incubations with rabbit renal cortical supernatant or microsomal suspensions based on mass spectroscopic data. We report herein their structure confirmation and synthesis in sufficient quantities for biological testing and biochemical study.

33





 $\underline{10}$: **R** = **H** ; **X** = **H**,OH 11: R = H; X = O



,O

Methyl 14,15-epoxyarachidonate⁶ (<u>1</u>) was hydrolyzed in 10% perchloric acidtetrahydrofuran (3:8) at 0° for 24 h to diol 2^7 (SiO₂:5% MeOH/CH₂Cl₂, R**f** ~0.43). Lead tetraacetate (1.05 equiv) cleavage in dry methylene chloride at -20°C for 15 min and warming to 0°C over 15 min afforded unstable aldehyde <u>3</u> (SiO₂:5% MeOH/CH₂Cl₂, R**f** ~0.59). The reaction mixture was filtered rapidly through a silica gel-Celite bed, evaporated with heptane and dried in vacuo. Wittig <u>cis</u>-olefination⁸ with phosphonium ylide <u>4</u>⁹ (1.5 equiv) in tetrahydrofuran-hexamethylphosphoric triamide (9:1) at -78°C for 30 min and warming to 0°C over 90 min afforded methyl ester <u>5</u> in 34% yield over-all from <u>1</u> [nmr (CDCl₃, **§**):1.15-2.24 (m,13H), 2.36 (t, J ~7Hz, 2H), 2.72-3.00 (m,6H), 3.70 (s, 3H), 3.68 (t, J ~7Hz, 2H), 5.20-5.56 (m,8H); tlc (SiO₂):ether/hexane 1:1, R**f** ~ 0.26]. Saponification of <u>5</u> produced <u>6</u> (SiO₂:5% MeOH/CH₂Cl₂, R**f** ~ 0.25) identical in all respects with the enzymatic product¹⁰. Jones oxidation of <u>5</u> at 0°C and saponification furnished <u>7</u> indistinguishable from natural material¹⁰.

Similarly, Wittig <u>cis</u>-olefination⁸ of <u>3</u> using ylide <u>8</u>¹¹ gave methyl ester <u>9</u> in 30% yield over-all from <u>1</u> [nmr (CDCl₃, **6**):1.23 (d, J ~7 Hz,3H), 1.38-2.24 (m,10H), 2.36 (t, J ~7 Hz,2H), 2.68-3.02 (m,6H), 3.68-3.74 (m,1H), 3.70 (s,3H), 5.20-5.56 (m,8H); tlc (S10₂):ether/hexane 1:1, R**f** ~ 0.22]. Saponification yielded <u>10</u> (Si0₂:5% MeOH/CH₂Cl₂, R**f** ~ 0.14). Jones oxidation and hydrolysis of <u>9</u> afforded keto-acid <u>11</u> [nmr(CDCl₃, **5**): 1.24-2.56 (m,12H), 2.18 (s,3H), 2.68-3.00 (m,6H), 5.20-5.56 (m,8H); tlc (Si0₂):ether/hexane 1:1, R**f** ~ 0.17]. Both <u>10</u> and <u>11</u> were identical with enzymatically derived material¹⁰. We are currently investigating the biological activity of the above described metabolites and the unprecedented¹² conversion by cytochrome P-450 of an unactivated methyl to carboxyl using synthetic material prepared from radiolabeled arachidonic acid.

<u>Acknowledgment</u>: This work was supported generously by the Robert A. Welch Foundation (I-782) and USPHS NIGMS-16488.

References and Notes

1.(a) J. Capdevila, N. Chacos, J. Werringloer, R.A. Prough, R.W. Estabrook, <u>Proc. Natl.</u>
<u>Acad. Sci. USA</u>, 78, 5362-5366 (1981); (b) J. Capdevila, L.J. Marnett, N. Chacos, R.A. Prough,
R.W. Estabrook, <u>ibid.</u>, 79, 767-770 (1982); (c) J. Capdevila, L. Parkhill, N. Chacos, R.
Okita, B.S.S. Masters, R.W. Estabrook, <u>Biochem. Biophys. Res. Comm.</u>, 101, 1357-1363 (1981).

A.R. Morrison, N. Pascoe, Proc. Natl. Acad. Sci. USA, 78, 7375-7378 (1981); E.H. Oliw,
 J.A. Lawson, A.R. Brash, J.A. Oates, J. Biol. Chem., 256, 9924-9931 (1981).

3. N. Chacos, J.R. Falck, C. Wixtrom, J. Capdevila, <u>Biochem. Biophys. Res. Comm.</u>, 104, 916-922 (1982).

4. J. Capdevila, N. Chacos, J.R. Falck, S. Manna, A. Negro-Vilar, S.R. Ojeda, submitted for publication.

5. J. Capdevila, N. Chacos, R.W. Estabrook, unpublished results.

6. E.J. Corey, H. Niwa, J.R. Falck, J. Amer. Chem. Soc., 101, 1586-1587 (1979).

7. Satisfactory infrared, proton magnetic resonance, and mass spectral data were obtained for all new compounds using chromatographically homogeneous samples, except for aldehyde $\underline{3}$ which was used without purification.

8. The olefin stereochemistry in <u>5</u> and <u>9</u> was confirmed by the sequence: tosylation, lithium aluminum hydride reduction, and Jones oxidation affording in both cases a single product (IR: 680 cm^{-1}) identical in all respects with arachidonic acid and distinguishable from eicosacis-5,8,11-trans-14-tetraenoic acid (IR:970 cm⁻¹) by TLC (AgNO₃/SiO₂). Trans isomer prepared from the bromo-mesylate of <u>1</u> using Zn/HOAc, E.J. Corey, A. Marfat, J.R. Falck, J.O. Albright, <u>J. Amer. Chem. Soc.</u>, 102, 1433-1435 (1980).

9. Prepared from 6-chlorohexanol by displacement with sodium iodide in refluxing acetone for 8 h, then triphenylphosphine/potassium carbonate in refluxing acetonitrile for 10 h, salt mp 136-137°C ($C_{6}H_{6}$). Ylide generation using two equivalents n-BuLi in THF at -78°C and warming to -30°C over 30 min.

10. Comparisons of enzymatic and synthetic products based on normal and reverse phase high pressure liquid chromatography (HPLC) and positive chemical ionization mass spectroscopy with methane as carrier gas. Consult ref. 2.

11. D.P. Curran, D. Scholz, Monatsh. Chem., 108, 1401-1411 (1977).

12. R.E. White, M.J. Coon, Ann. Rev. Biochem., 49, 315 (1980).

(Received in USA 13 July 1982)